

## 4-HYDROXY-*N*-METHYLPROLINE ANALOGUES IN *MELALEUCA* SPP.

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**Key Word Index**—*Melaleuca*; Myrtaceae; (–)-(2*S*, 4*R*)-1-methyl-4-hydroxy-pyrrolidine-2-carboxylic acid; 4-hydroxy-*N*-methylproline; (–)-(2*S*, 4*R*)-1,1'-dimethyl-4-hydroxypyrrolidine-2-carboxylic acid; 4-hydroxy-*N,N'*-dimethylproline; betonicine.

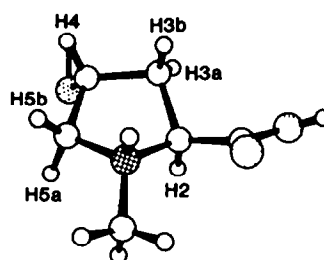
**Abstract**—4-hydroxy-*N*-methylproline and 4-hydroxy-*N,N'*-dimethylproline have been identified in the aqueous extracts of *Melaleuca* spp. Characterization of the two compounds is discussed.

Whilst using  $^1\text{H}$  NMR to measure the accumulation of glycinebetaine in a range of native South Australia flora [1] it was observed that a specimen of *Melaleuca lanceolata* contained a high concentration of an *N*-methyl hydroxyproline derivative. Subsequent studies have shown that a number of species belonging to the genus *Melaleuca* accumulated this compound and another hydroxyproline analogue. The levels of these compounds in plant tissue respond to environmental stress (Naidu, B. P., Jones, G. P., Pales, L. G. and Poljakoff-Mayber, A., unpublished data) suggesting that they may be important in determining the drought and/or salinity resistance of these species. Furthermore, the occurrence of these compounds together with proline and *N*-methylproline, at different levels in different *Melaleuca* species holds promise for their use in chemotaxonomic studies. The compounds were isolated by ion-exchange chromatography and identified as (–)-(2*S*, 4*R*)-1-methyl-4-hydroxy-pyrrolidine-2-carboxylic acid (4-hydroxy-*N*-methylproline, 1) and (–)-(2*S*, 4*R*)-1,1'-dimethyl-4-hydroxy-pyrrolidine-2-carboxylic acid (4-hydroxy-*N,N'*-dimethylproline, 2) by  $^1\text{H}$  and  $^{13}\text{C}$  NMR and by comparison with authentic compounds. X-ray crystallographic analyses of the two compounds enabled unambiguous confirmation of their identities. The conformations of 1 and 2 determined by X-ray diffraction studies of their respective hydrochlorides are shown below. Oxygen and nitrogen atoms are lightly and heavily shaded respectively. The orientation of the *N*-methyl group with respect to the carboxyl group in 1 is the same as that predicted by an earlier  $^1\text{H}$  NMR study [5].

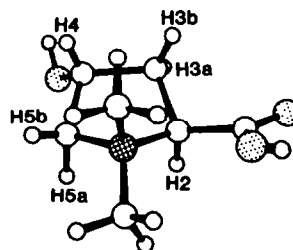
1 and 2 have both been found in higher plants [2–4] and algae [5–7] at levels less than 0.5% dry weight. In the present case levels of 1 of >4% dry weight were measured in *M. lanceolata* and >1.4% dry weight of both 1 and 2 in leaves of *M. uncinata*.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectral data for 1 are consistent with that reported previously [5]. The  $^1\text{H}$  NMR spectrum of 2, as the hydrochloride (90 MHz,  $\text{D}_2\text{O}$ , pH 1.0), closely resembled that of 1 and showed peaks at  $\delta 3.19$  (3H, *N*-Me) and  $\delta 3.47$  (3H, *N'*-Me). A complex multiplet at  $\delta 2.25$ – $3.00$  (2H) was attributed to H-3a and H-3b. H-5a and H-5b each gave a doublet of doublets centred at  $\delta 3.62$  and  $\delta 4.11$  respectively as part of an AMX pattern involving H-4 where  $J_{5a,5b} = 13.0$  Hz,

$J_{3a,4} = 4.4$  Hz and  $J_{5b,4} = 6.0$  Hz. A multiplet at approximately  $\delta 4.7$ , which was significantly obscured by the solvent peak, was ascribed to H-2 and H-4. When the pH was adjusted to 4, a part of this multiplet moved upfield to form a broad doublet at  $\delta 4.36$  (1H),  $J = 11$  Hz which was ascribed to H-2.

The  $^{13}\text{C}$  NMR spectrum of 2 (22.5 MHz,  $\text{D}_2\text{O}$ , pH 1.0) shows a singlet at  $\delta 171.0$  (–COOH), two doublets at  $\delta 77.5$  and  $\delta 76.8$  (C-2, C-4), two triplets at  $\delta 69.1$  (C-5) and  $\delta 38.1$  (C-3) and two quartets at  $\delta 57.6$  and  $\delta 52.0$  (*N*-Me, *N'*-Me). At pH 4.0 the carboxyl signal moved downfield to  $\delta 173.0$  and the signals associated with C-2 and C-4 merged to form a single doublet at  $\delta 76.9$ .



1



2

It is noted that the melting points of the hydrochlorides for both 1 and 2 are significantly lower than those reported previously [2, 5] and this is attributed to the formation of different crystalline modifications.

#### EXPERIMENTAL

**Extraction and isolation.** Leaves of *M. lanceolata* (450 g fr. wt) were homogenized and extracted with 70% aq MeOH (3 × 1). The extracts were filtered through muslin and combined. The filtrate was then clarified by centrifugation, concd *in vacuo* and again centrifuged. The supernatant was applied to a column of Dowex 50W (H<sup>+</sup> form, 50–100 mesh, column size 3 × 65 cm) and after washing the resin with water (2l) to remove neutral species, the amino acid fraction containing 1 was eluted with 4 M HCl (1l). The eluent was taken to dryness *in vacuo*. The reddish-brown solid was taken up in hot dry EtOH (60 ml) to which was added dry Et<sub>2</sub>O to ppt. the hydrochloride of 1 as pale pink crystals (yield 7.5 g). A small sample of this material (200 mg) was recrystallized from MeOH–Et<sub>2</sub>O to give colourless prisms suitable for X-ray crystallographic studies with mp 160–161° (lit. 183–184° (dec) [5]),  $[\alpha]_D^{25} - 59.8^\circ$  (H<sub>2</sub>O; c 1.1) (lit.  $- 58.9^\circ$ ; H<sub>2</sub>O; c 1.2) [5]. A sample of the hydrochloride of 1 (200 mg) was converted to the free amino acid on a Dowex 50 W [H]<sup>+</sup> column by elution with 2 M NH<sub>4</sub>OH. The elute was dried *in vacuo* and the colourless residual solid was recrystallized from MeOH–Et<sub>2</sub>O giving 1 as colourless needles (120 mg), mp 239° (dec) (lit. 238° (dec) [5]),  $[\alpha]_D^{25} - 82.2^\circ$  (H<sub>2</sub>O; c 1.0) (lit.  $- 82.7^\circ$ ; H<sub>2</sub>O; c 1.1 [5]), FAB-MS (M + H)<sup>+</sup> 146.

*M. uncinata* leaf tissue (350 g) was extracted in a similar manner. After the initial ion exchange chromatography stage, however, 1 and 2 were sepd on a second Dowex 50W column (50–100 mesh; H<sup>+</sup>, 3 × 65 cm) with a stepwise gradient of HCl from 0 to 1 M in 0.3 M steps (2l) collecting 100 ml fractions. The

separation was monitored by TLC analysis (precoated silica gel plates using EtOH–conc NH<sub>4</sub>OH, 85:15) using iodine and Dragendorff's reagent for visualizing the spots. 1 was eluted at the end of the 0.6 M HCl step whereas 2 was eluted with 0.9 M HCl. Fractions containing either 1 or 2 were pooled, dried *in vacuo* and recrystallized from MeOH–Et<sub>2</sub>O yielding 650 mg of 1 and 620 mg of 2 as the hydrochlorides. The physical properties of 1 were the same as the material isolated from *M. lanceolata*. Crystals of 2 as the hydrochloride, suitable for X-ray crystallographic analysis (X-ray crystallographic data for the two compounds have been deposited with the Cambridge Crystallographic Data Centre.) are colourless prisms, mp 204–205° (lit 224° (dec) [2]),  $[\alpha]_D^{25} - 26.4^\circ$  (H<sub>2</sub>O; c 1.1) (lit.  $- 24.8^\circ$ ; H<sub>2</sub>O; c 3.6 [2]), FABMS (M + H)<sup>+</sup> 160.

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#### REFERENCES

1. Poljakoff-Mayber, A., Symon, D. E., Jones, G. P., Naidu, B. P. and Paleg, L. G. (1987) *Aust. J. Plant Physiol.* (in press)
2. Goodson, J. A. and Clewer, H. W. B. (1919) *J. Chem. Soc.* 923.
3. Delaveau, P., Koudogbo, B. and Pousset, J. L. (1973) *Phytochemistry* 12, 2893.
4. Morgan, J. W. W. (1964) *Chem. Ind.* 542.
5. Sciuto, S., Chillemi, R., Piatelli, M. and Impellizzeri, G. (1983) *Phytochemistry* 22, 2311.
6. Blunden, G., Gordon, S. M., McLean, W. F. H. and Guiry, M. D. (1982) *Bot. Marina* 25, 256.
7. Hori, K., Yamamoto, T., Miyazawa, K. and Ito, K. (1979) *Hiroshima Daigaku Seibutsu Kiyo* 18, 65.